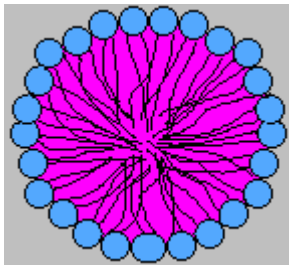
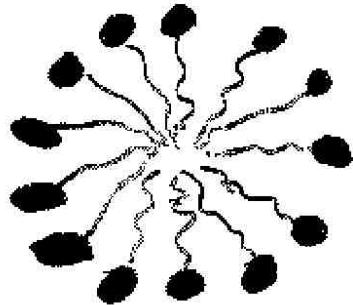


Lipid Self-assembly

Self-assembly occurs due to the thermodynamics, if the phospholipids are in water (or other polar solution) the tails will want to be 'away' from the solution. They could all go to the top (like oil on water), or they could have the tails point toward each other. With the tails pointing toward each other, this could result in 2 different forms.

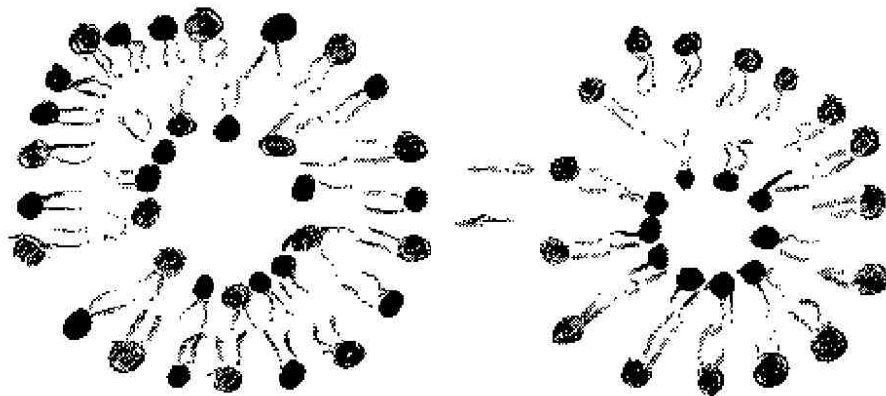
First would be a [micelle](#) which would like a ball with the phospholipid heads on the outside and the tails pointing together like this or in the form of a lipid bilayer:





Micelle
(Single chain)

Mixture of lipids in bilayers



homogeneous

segregated

Possibility of lateral phase separation.

Micelles

Are made of single chain amphiphiles

Are aggregates with polar head groups exposed to the surface in contact with water and the hydrocarbon portion clumped together.

Micelle interior

Similar to pure hydrocarbon solution
Comparison of free energy of transfer

Water to micelle interior				Water to liquid hydrocarbon		
	ΔG	ΔH	ΔS	ΔG	ΔH	ΔS
Ethane	-3.45	+2.0	18.3	-3.9	+2.5	21
Propane	-4.23	+1.0	17.5	-4.9	+1.7	22
Butane	-5.13	+0.0	17.2	-5.9	+0.8	23

These data indicate that the micelle interior is similar to hydrocarbon

The size and the shape of a micelle depends on the ratio of the surface area

$$A_s$$

to the number of head groups

$$N_h$$

This is due to the importance of repulsive component. As the size of the micelle increases the ratio A_s/N_h decreases.

If l is the radius of a micelle, we can assume

$$l = aN_c = a'N_h$$

for single chain amphiphiles,

N_c is the number of hydrocarbon atoms per chain.

$$A_s = 4\pi l^2 = 4\pi a^2 (N_c)^2 \quad (\text{sphere})$$

$$V = 4/3\pi r^3 = 4/3\pi a^3 (N_c)^3$$

$$A_s/N_h = 3b/a$$

For a cylinder

$$A_s/N_h = 2b/a$$

For a large planar bilayer

$$A_s/N_h = b/a$$

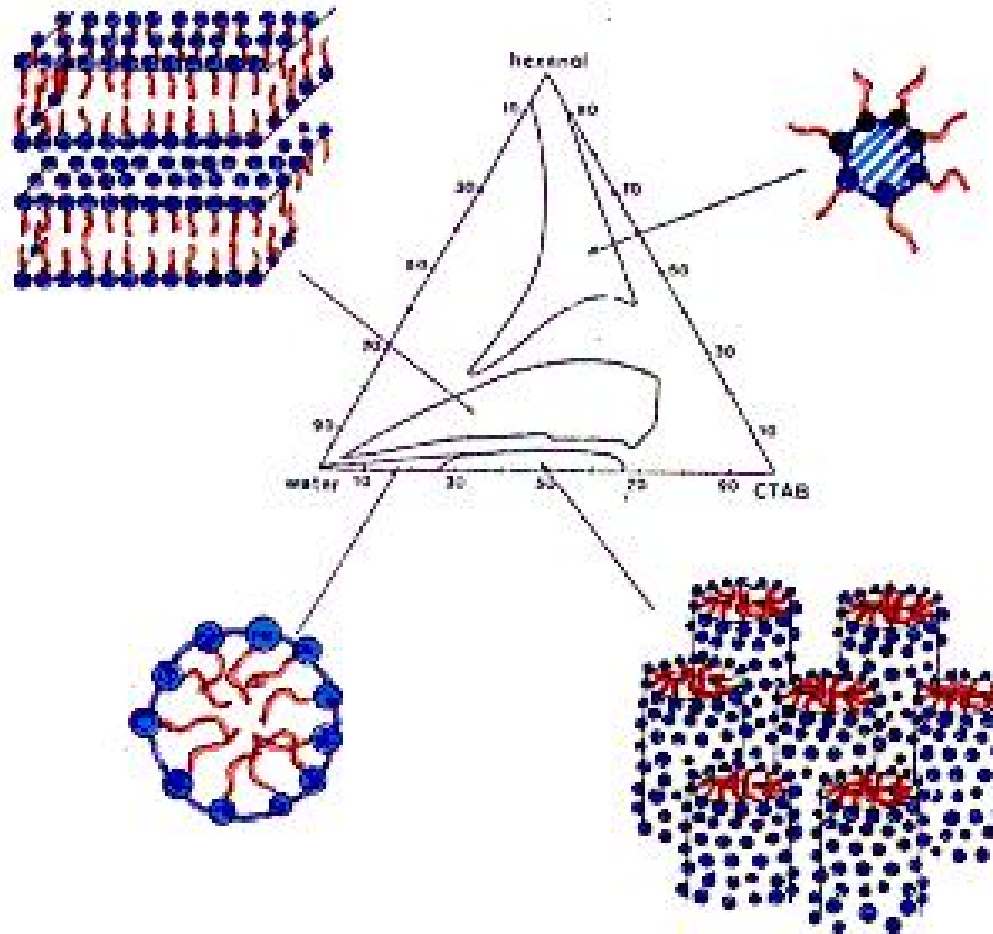
The larger the number of molecules per micelle, the more planar the structure of the micelle will be.

Reverse Micelles

In some organic solvents, amphiphiles form a micelle in which the charged groups are in the interior.

Driving force? Some water in the micelle interior

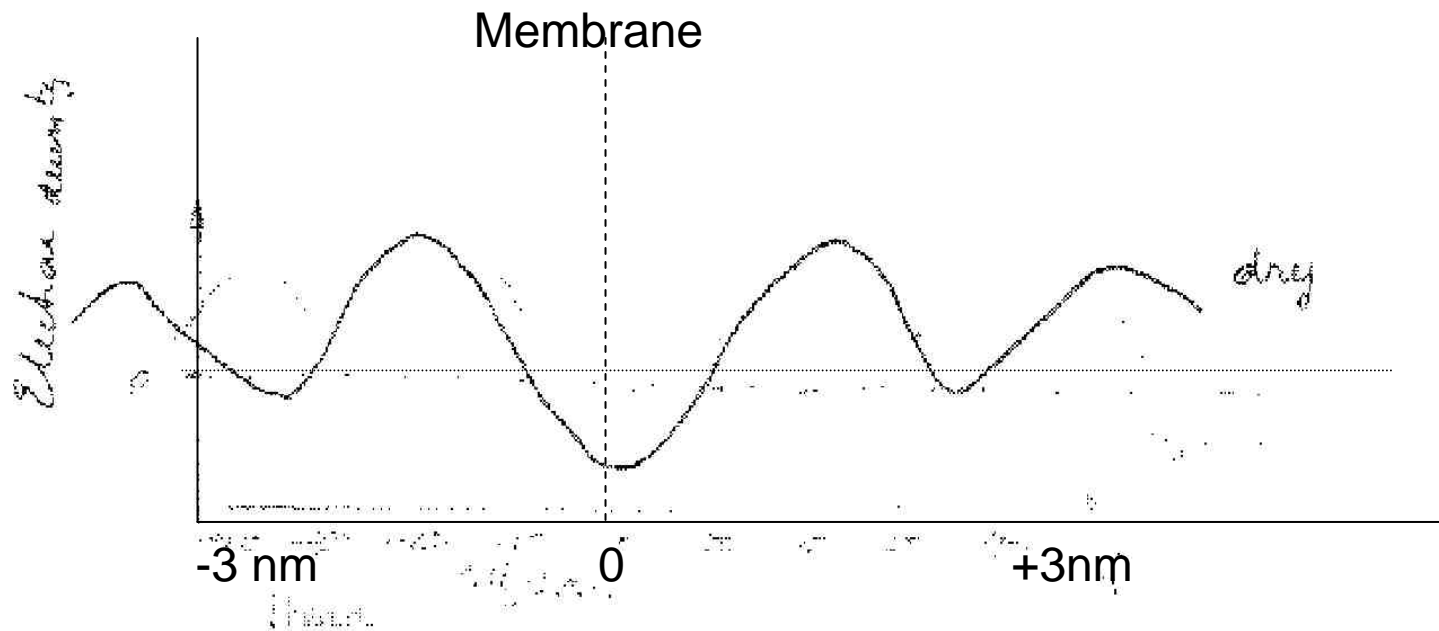
Micelles and Microemulsions



Amphiphilic molecules spontaneously self-assemble in solution to form a variety of aggregates. In our research we focus on two main topics: (i) the use of surfactant solutions as interesting and versatile model systems in polymer and colloid physics (micelles as equilibrium polymers and polyelectrolytes); and (ii) on the various non-equilibrium or metastable states and the pathway and kinetics associated with structural transitions and phase separation.

Bilayers

Structural and dynamical features of bilayers

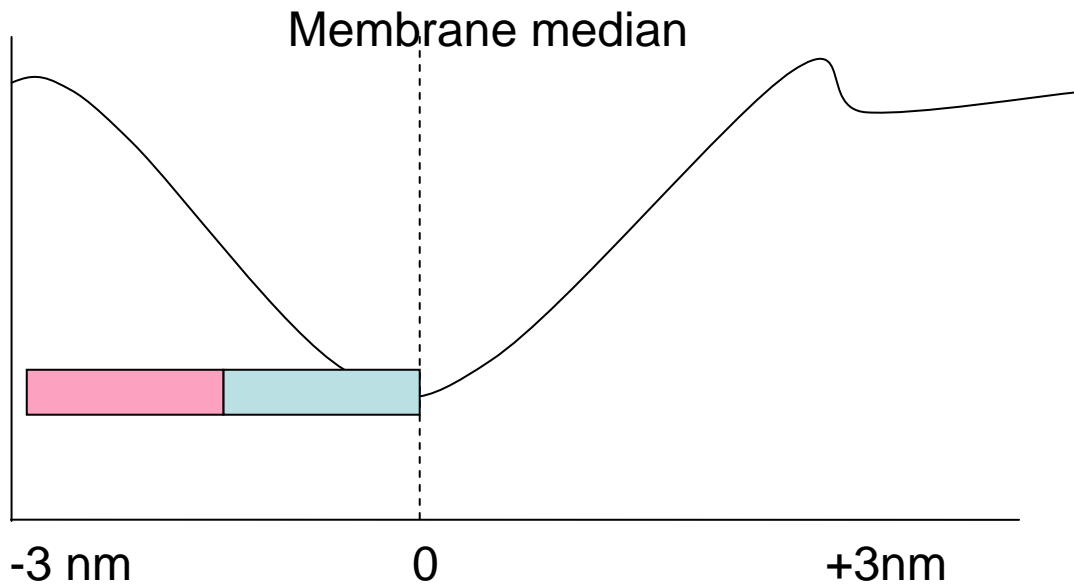


(from x-ray diffraction)

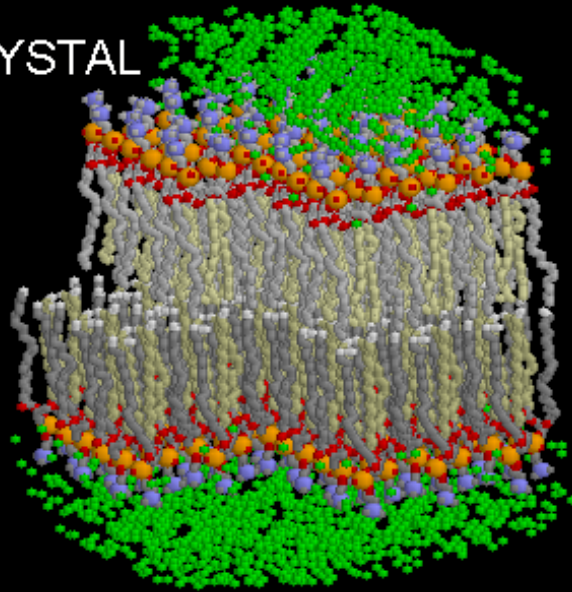
The minimum in electron density is due to CH_3 groups. They have the lowest density.

The hydrocarbon chains are not interdigitated. They are oriented perpendicular to the layer plane. (Effect of cholesterol)

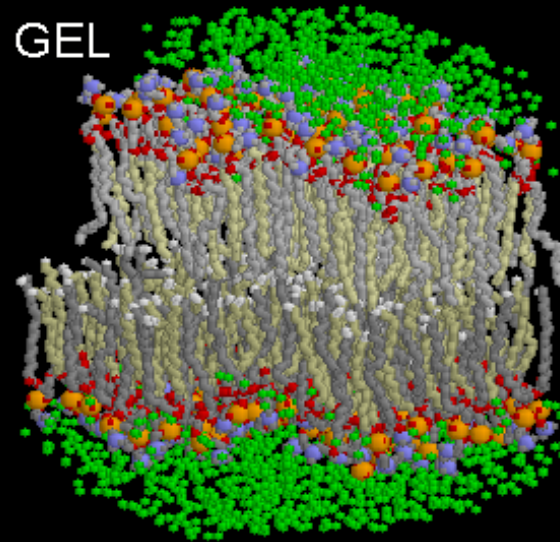
Penetration of water (from EPR studies): Plot of polarity index Water molecules penetrate a lot inside the bilayer



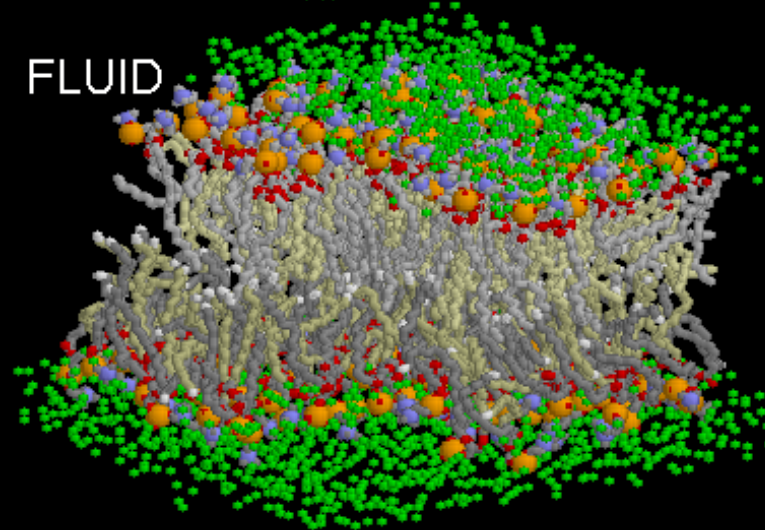
CRYSTAL



GEL



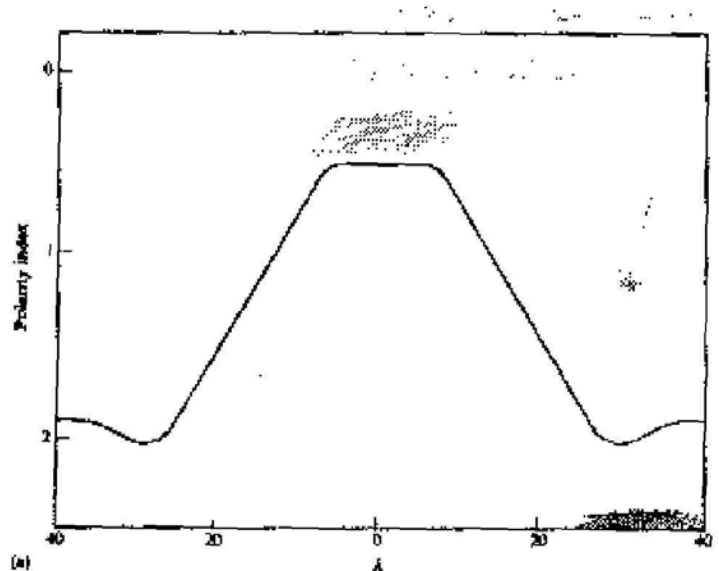
FLUID



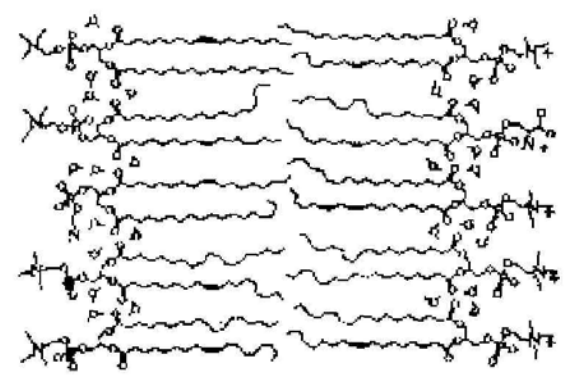
Molecular Dynamics Simulation
of Phosphatidyl Choline Bilayer

Carbon/Palmitic Oleic
Nitrogen Oxygen Phosphorus
Water Oxygens

H Heller, M Schaefer, K Schulten,
J Phys Chem 97:8343, 1993.
RasMol Image by E Martz



(a)



(b)

Figure 28-28
 The hydrophobic barrier in a lipid bilayer. (a) Polarity index across the bilayer. (b) A schematic view of the bilayer. [After O. H. Griffith et al., *J. Membrane Biol.* 15:159 (1974).]

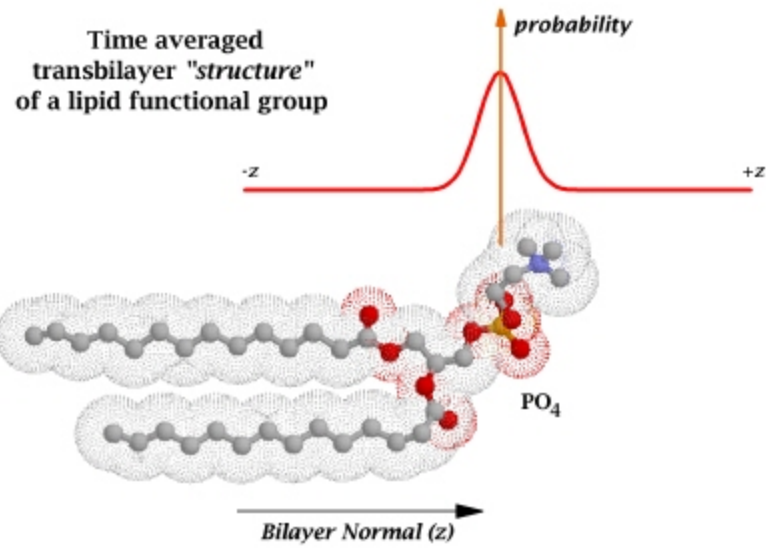
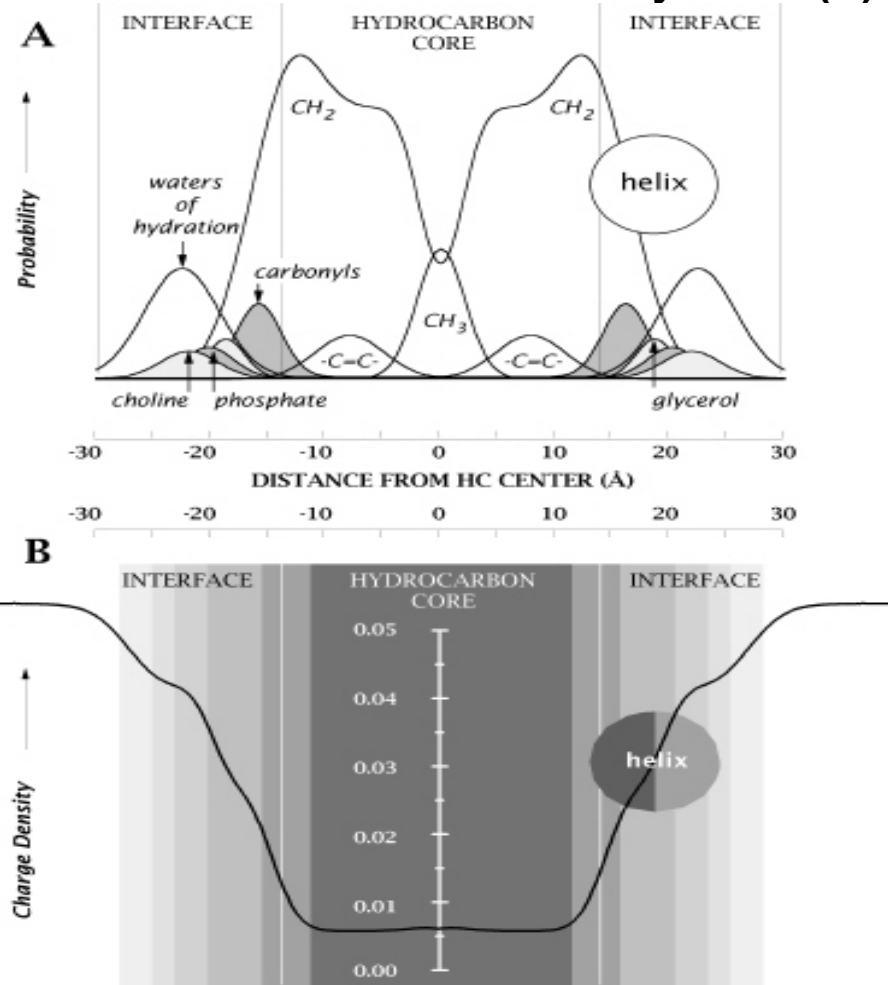
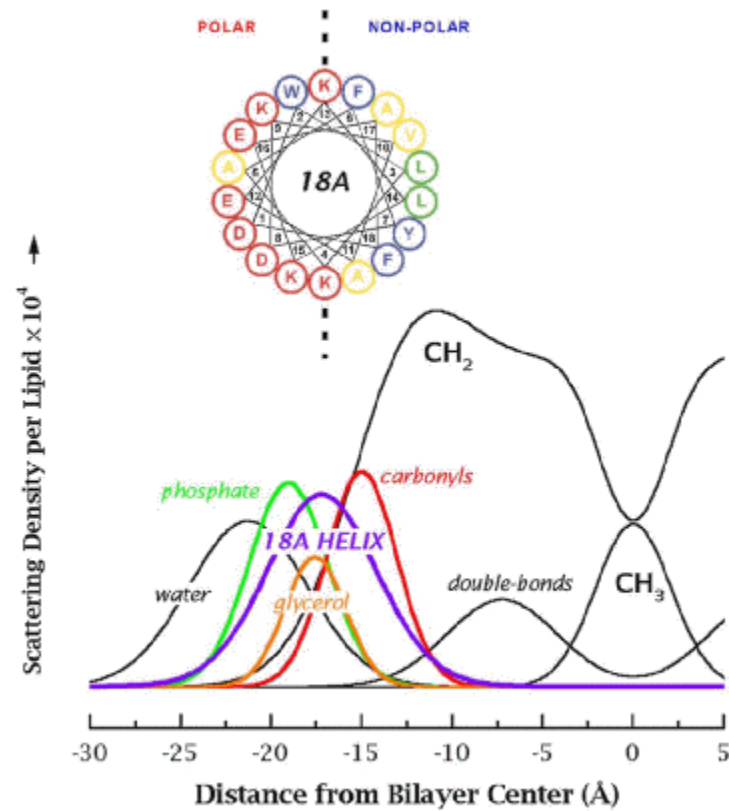


Figure 2. (A) The Structure of a Fluid DOPC Bilayer and (B) Its Polarity Profile (go to



[Fig. 1](#) or [Fig. 3](#))

Figure 3. An Amphipathic Alpha-Helix in the Fluid DOPC Bilayer (go to [Fig. 1](#) or [Fig. 2](#))



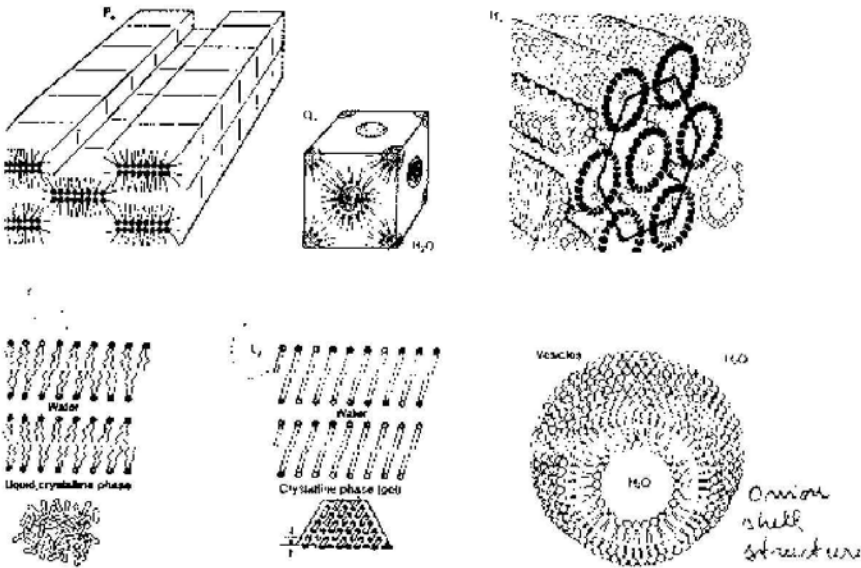
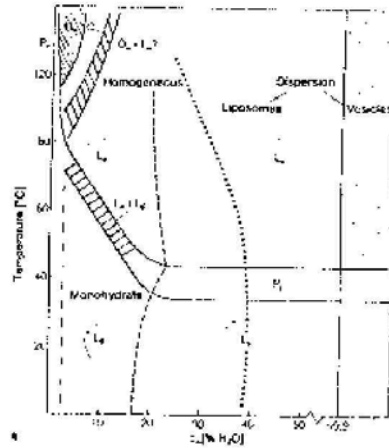
Flip-flop transitions and lateral diffusion

F-F are very rare

lateral diffusion is relatively fast (EPR) $1.8\mu\text{m}^2/\text{s}$

Bilayers undergo a thermal transition between a low temperature gel-like ordered state and a high temperature liquid-crystalline state

Fig. 12.7. Phase diagram of dipalmitoylphosphatidylcholine (DPPC) in water. The abscissa is the concentration of water, with pure DPPC at the left. The lamellar phases L_1 and L_2 are liquid and L_c is crystalline phases in which the lipid chains form a regular lattice. *Horizontal* curves show the maximal water absorption of a vapor, and *vertical* curves show the corresponding absorption from a liquid. To the right of the latter curve the lipid-water system is aqueous. Below 2.5%, water there could be in existence between hydrate and water free, crystalline lipid. The shaded regions of coexistence between L_1 and L_c , and between L_2 and L_c , and at high temperatures, there are several liquid crystalline states, such as the hexagonal phase H_2 , the cubic phase of the ribbon phase F_2 , whose structure is illustrated schematically (Pouyet and Pershan, Biophys. J. 20: 117 (1972); Comparison of L. in Phys. Jap. 1: 445 (1971); Arigona, B. Dissertation, University of Lausanne, 1981).



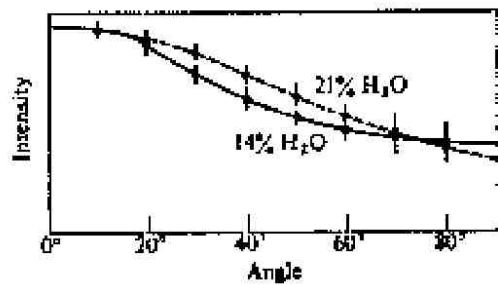


Figure 25-11

*Angular variation of intensity of the hydrocarbon-chain band of egg lecithin bilayers. The radially integrated intensity of the 4.6 Å diffraction band is shown from the equator (0°) to the meridian (90°). [After Y. K. Levine and M. H. F. Wilkins, *Nature New Biology* 230:69 (1971).]*

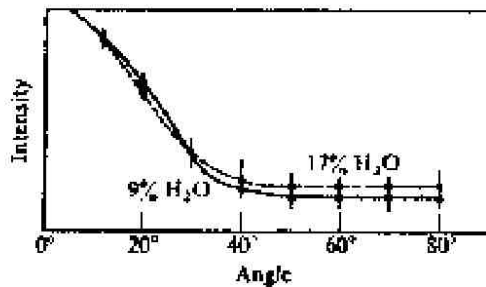


Figure 25-12

*Angular variation of intensity of the hydrocarbon-chain band of egg lecithin bilayers in the presence of cholesterol. The radially integrated intensity of the 4.75 Å diffraction band is shown from the equator (0°) to the meridian (90°). [After Y. K. Levine and M. H. F. Wilkins, *Nature New Biology* 230:69 (1971).]*

Biological membranes

The fluid mosaic model (Singer)

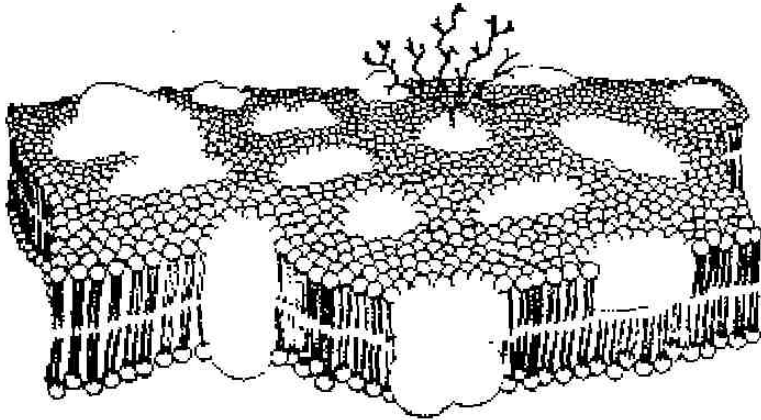


Figure 4-12

A general model for the structure of biological membranes. [After J. M. Clark, Jr. and R. L. Smitzer, *Experimental Biochemistry*, 2nd ed. (San Francisco: W. H. Freeman and Company, Copyright © 1977).]

Composition

	Protein	Lipid	Carbohydrate
Myelin	18%	79%	3%
Erythrocyte	48%	43%	8%
Chloroplast	70%	30%	(2%)
Mitochondrial	76%	24%	(1-2%)
Purple membrane	75%	25%	

Protein-lipid interactions

Lipids

Proteins

Hydrophobic
tail

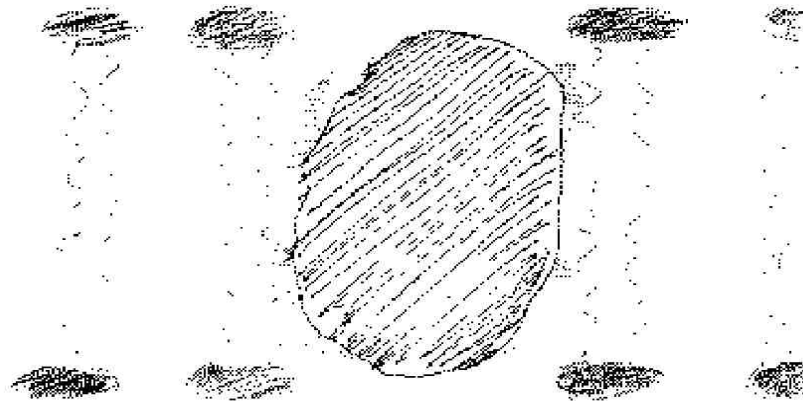
Non-polar aminoacid
Side chains

Polar heads

Polar aminoacids

Protein -lipid interface

Perturbed state
(similar to hydrocarbons?)

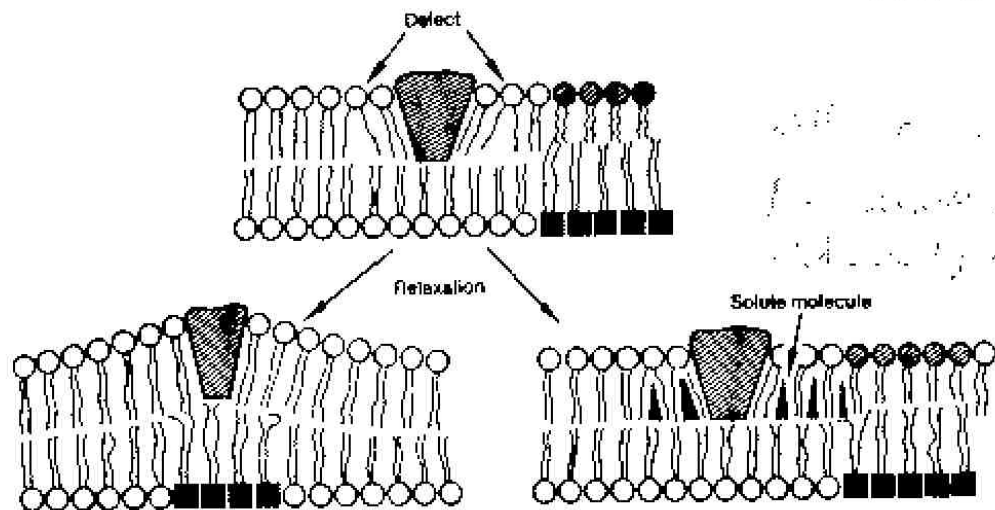


- Recognition of a particular lipid
- Penetration of lipids into the protein

Arrangement of Membrane proteins

Distribution on the membrane surface





with a conical protein
 for example of a
 cholesterol

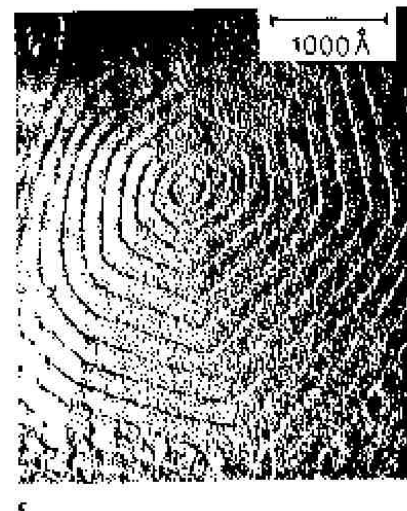
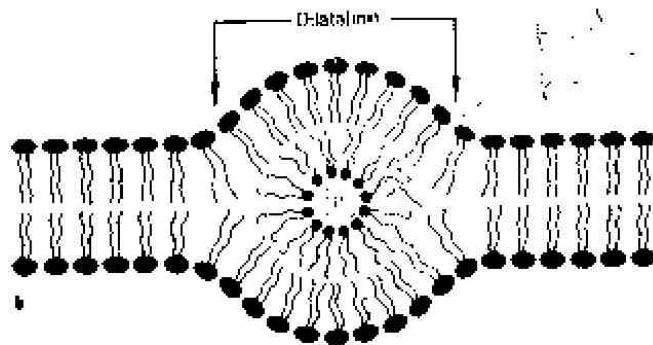
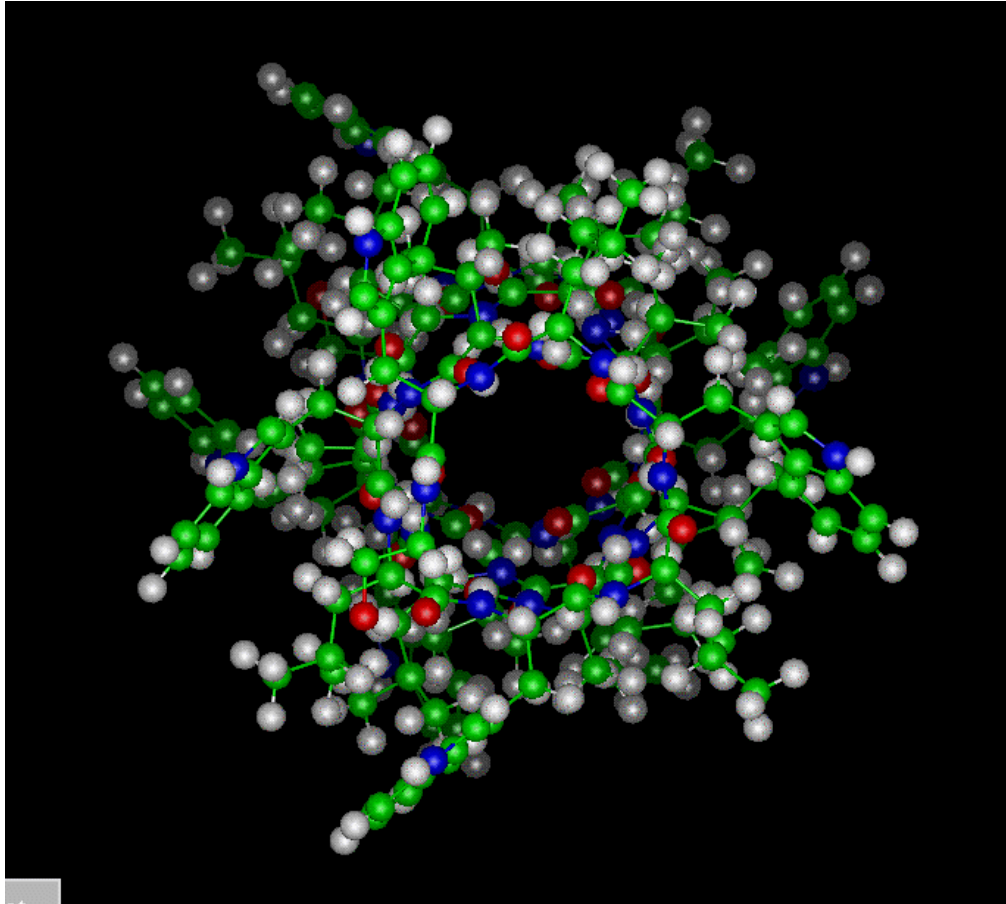


Fig. 12.21. a Schematic representation of two possible mechanisms for relaxation of elastic stress at orientation defects around a conical protein. *Left*: escape into the third dimension and subsequent redistribution of lipids in the lower monolayer. *Right*: incorporation of a swarm of foreign molecules. b Schematic representation of the intercalation of a protein between monolayers. Protein is enclosed in an inverted micelle. The intercalation leads to the formation of a dilution region inside the bilayer. c Electron microscope image of a freeze-etched DMPC bilayer below the main transition showing a defect

structure which has been stabilized by the incorporation of 4 mol % cholesterol (cf. Fig. 12.8a where the defect line forms a spiral rather than closed hexagons)



Gramicidin channel

Aquaporin Channel



Peter Agre

